

## Monday, October 21st, 2019 4:00-5:00 PM BSRB, ABC Seminar rooms



## "RNA Editing Enzyme Machines"

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## Abstract

Parasitic protist *Trypanosoma brucei* causes African human and animal trypanosomiasis, a spectrum of diseases affecting the population and economy in sub-Saharan Africa. These digenetic hemoflagellates belong to *Kinetoplastea*, a taxonomic class distinguished by possession of a kinetoplast. This nucleoprotein body contains mitochondrial DNA of two kinds: ~25 maxicircles (each ~23kb) encoding ribosomal RNAs, two guide RNA (gRNAs), ribosomal proteins and subunits of respiratory complexes, and approximately 5000 of ~1kb minicircles bearing the majority of gRNA genes. Relaxed maxicircles and minicircles are interlinked and packed into a dense disc-shaped network by association with histone-like proteins. Both maxicircle and minicircle genomes are transcribed by a phage-like RNA polymerase from multiple promoters into 3'-extended precursors which undergo 3'-5' exonucleolytic trimming. To function in mitochondrial translation, pre-mRNAs must further proceed through 3' adenylation, and often gRNA-directed uridine insertion/deletion editing, and 3' A/U-tailing. Ribosomal and guide RNAs are typically 3' uridylated. Historically, the fascinating phenomenon of RNA editing has attracted major research efforts, but more recent developments provided insights into pre- and post-edited processing events and identified key players in transforming primary precursors into functional RNAs and regulating their turnover. I will present a forward-looking model that integrates known modalities of mitochondrial RNA metabolism.